DIFFERENTIAL EXPRESSION OF MICRORNAS IN LUNG CANCER: A REVIEW ARTICLE

Omair Ahmad, Zakiya Saiyada, Mohammad Armoghan, Bilal Khan, Swapnil Raj Gautam, Zainab Siddiqui*

Research Metabolic Unit, Department of Pathology*

Era's Lucknow Medical College & Hospital, Era University, Sarfarazganj, Lucknow, U. P., India - 226003

ABSTRACT

MicroRNAs (miRNAs) are small non-coding RNA molecules of approximately 16-24 nucleotide length. The miRNA biogenesis is a 2 step cleavage process mediated by Dorsha and Dicer. The nuclear cleavage by Dorsha / DiGeorge syndrome critical region 8 (DGCR8) generates 60-70 nucleotide long precursor microRNA (pre-miRNA). Furthermore, the pre-miRNA is exported to the cytoplasm by exportin 5 to be cleaved by Dicer. This resultant miRNA is further processed to generate a mature miRNA and get assembled into a RNA-induced silencing complex (RISC). Hence leading to transcriptional repression of the target mRNAs. It has been reported that one miRNA may target Received on : 17-02-2022 Accepted on : 11-05-2022

Address for correspondence **Dr. Zainab Siddiqui** Department of Pathology Era's Lucknow Medical College & Hospital, Era University, Lucknow, 226003 Email: zainab.siddiqui6@gmail.com Contact no: +91-8299350733

many genes accounting from a few to as many as thousands. Lung cancer (LC) ranks third worldwide and is marked by poor prognosis. The early staged LC patients usually exhibit no symptoms and the condition worsens till the time of first diagnosis. Therefore, studies are required to outline good early detecting and surveillance biomarkers for LC. Several evidences support the role of miRNAs in the pathogenesis of LC. They show differential expression pattern i.e. may be either upregulated or downregulated. The oncogenic miRNAs remain upregulated while the tummor suppressive miRNAs remain downregulated. In LC miRNAs are the important factors for tumour initiation, differentiation, apoptosis, proliferation as well as tumor progression. Thus, this review article focuses on the diagnostic significance of miRNAs in LC.

KEYWORDS: Lung Cancer, microRNA, Oncogene, Tumor Suppressor, Diagnosis

INTRODUCTION

LC is one of the most common leading cause of cancer related deaths globally. It ranks 3rd amongst all the cancers preceded by breast and prostate cancer. According to The Global Cancer Observatory (GLOBOCAN)-2018 data, the estimated agestandardized incidence rates (for both sexes) for LC was 11.6 %. While the mortality rate was found to be 18.4%, thus accounting for the highest number of deaths in 2018 (1). It has also been predicted that males have higher rate of mortality than females i.e. 22.0% and 13.8% respectively. The rate of diagnosis is also higher for males (14.5%) than females (8.4%) (2). Since LC is associated with high mortality rates it is important to evaluate the significance of early diagnostic biomarker. Early staged LC patients usually experience no symptoms and the disease remains undiagnosed for a longer period of time (3, 4).

On the contrary, higher LC stages are difficult to treat and are associated with low survival rate. According to GLOBOCAN 2014 data, the survival rate of LC patient was lowest 14.70% (5-year net survival) in the United Kingdom and the highest survival rate was observed in Canada 22.60% (5).

Several human studies have shown promising results for the evaluation of miRNAs as diagnostic biomarkers in LC (6, 7, 8). The miRNA was first discovered in 1993 by Ambros and colleagues in Caenorhabditis elegans. It is a small non-coding RNA molecule of about 22 nucleotides. They are known to regulate the gene expression post-transcriptionally (9). miRNAs can be oncomir or tumour suppressors depending upon the nature of their activity (10). OncomiRs are cancer causing and are found to be upregulated in LC patients such as miRNA-17, miRNA-21, miRNA-155 and miRNA-27a(6, 7, 11,12) while tumor suppressors control the abnormal growth of cells by inhibiting various oncogenic factors. Few tumour suppressors such as miRNA-126, miRNA-let-7a, miRNA-29b-3p and miRNA-145 are reported to be down regulated in LC patients (13, 14, 15, 16).

MICRORNABIOGENESIS

The miRNAs are produced endogenously in our body through a regulated process. The double stranded miRNA is composed of a single stranded RNA complementarity paired with its own nucleotide. This results in a hairpin loop like structure known as primary miRNA (primiRNA). The miRNA biogenesis is a two-step cleavage process performed firstly in nucleus then in cytoplasm (figure 1). The nuclear enzymes dorsha and RNA binding protein DGCR8 combine together to cut the pri-miRNA into small segments called pre-miRNA. Once the premiRNA is produced, it is transported to the cytoplasm via transporter protein exportin 5. The second cleavage process is mediated by an enzyme known as dicer (RNAse III type endonuclease) (17). It binds to the premiRNA and cuts it from two locations on the either ends. The resultant product is without a hairpin loop. This structure is known as miRNA: miRNA*duplex which is loaded on RISC having 'Slicer/TRNC6 and Argonaut' protein. Argonaut protein have two domains, PIWI and PAZ (18). This double stranded RNA loaded on RISC will no longer be active unless one strand is released, which is a key task of RISC Complex (19, 20). One strand remain bound to RISC through PAZ domain and is known as the guide strand, while the other strand eventually leaves the RISC with the help of PIWI domain is known as the passenger strand. The guide strand binds complementarity to the target mRNA sequence. This process is called nucleation which represses translational activity by Argonaut protein having RNase H activity and cleaves the target mRNA to repress its translational activity. Guide strand with Argonaut protein are again ready to bind with another target mRNA (21)

CHARACTERISTICS OF miRNAs AS DIAGNOSTIC BIOMARKERS IN LUNG CANCER

miRNAs proves to be a potential diagnostic biomarker for early stage LC. The non-coding RNAs i.e. miRNAs has been long established as diagnostic and prognostic biomarkers in various type of cancers (18). In table 1, we have reported 42 miRNAs identified through extensive search of reported data in LC cases. The patients' characteristics are summarized in table 2. Generally, the expression value of significant miRNAs can either be low or high due to its association with the clinical characteristics of the patients while the expression value of non-significant miRNAs are not dependent on the patient characteristics.

In this review we have focused on the potential aspects of the various types of miRNAs studied as diagnostic biomarkers in LC patients.

Oncogenic miRNAs Targets/Functions in Lung Cancer

Shan et al., 2018 investigated targets of the miRNA-21-5p by DIANA-miRPath v3.0, a pathway analysis Web server. And further analysed the cascades regulated by them through KEGG database (26). The miRNA-21-5p was found to significantly regulate lysine degradation, fatty acid metabolism, adheren junctions, along with FoxO, Hippo, p53, prolactin, thyroid hormone, HIF-1, ErbB, and PI3K-AKT signalling pathways (26). The KEGG analysis also identified role of miRNA-21-5p in various other cancers such as renal cell carcinoma, prostate cancer,

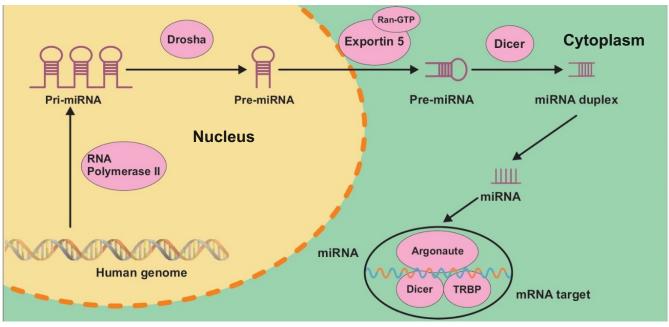


Fig. 1: Figure 1 miRNA Biogenesis Pictorial Representation

pancreatic cancer, endometrial cancer etc. (26). In another study researchers identified 709 target genes of miRNA-210 with the use of 12 online prediction software tools (30). Ten gene ontology terms and KEGG pathways were found by using DAVID 6.7. The gene ontology terms fell into the categories of molecular function, cellular component, and biological process. The highly significant pathways reported in KEGG were the cGMP-PKG signalling, renin secretion, and cell adhesion molecules (30). The authors also employed STRINGS to further narrow down the target genes of miRNA-210. They found 6 key miRNA-210 target genes i.e. cholinergic receptor muscarinic 2, adenylate cyclase 9, CXCL12, IL-6, G protein subunit γ 11, and adrenoceptor beta-2 (30).

Tumor Suppressive miRNAs Targets/Functions in Lung Cancer

The miRNA-let-7a mimic mediates the downregulation of dicer enzyme in lung cancer cell line (14). It also arrests lung cancer cells in G0/G1 phase and to a lesser extent in S phase, thus limiting cell proliferation (14). The authors also determined miRNA-let-7a prognostic significance and proved it to be an independent predictor of LC (14). Li and co-workers investigated the molecular pathways targeted by miRNA-195-5p (35). They retrieved nine significant signalling cascades through Ingenuity Pathway Analysis and suggested IL-8 pathway to be the most significant (35).

| S. No | S. No miRNA Type of sample | | Expression | AUC (95% CI) | Diagnostic value | Ref. No. | |
|-------|----------------------------|--------|-------------------|--|--|----------|--|
| 1 | miRNA- 33a-5p | Blood | Down regulated | | | 22 | |
| 2 | miRNA- 28-3p | Serum | Upregulated | 1.51 (1.24 – 1.85) | | 23 | |
| 3 | miRNA- 128-3p | Blood | Down regulated | 0.927 (0.861 – 0.993) | | | |
| 4 | miRNA- 17 | Plasma | Down regulated | 0.615 (0.574 – 0.655) | | 23 | |
| | | Plasma | Upregulated | 0.833 (0.577 – 0.966) | sensitivity =77.78% specificity =87.50% | 11 | |
| | miRNA- 17-5p | Serum | Upregulated | 0.738 (0.649 – 0.814) | sensitivity = 66.7% specificity = 76.6% | 24 | |
| 5 | miRNA- 222 | Plasma | Upregulated | 0.542 (0.289 – 0.780) | sensitivity = 50% specificity =88.89% | 11 | |
| 6 | miRNA- 190b | Plasma | Down regulated | 0.814 (0.779 – 0.845) | | 23 | |
| 7 | miRNA- 155 | Plasma | Upregulated | 0.8648 (0.8011–0.9329) | | 25 | |
| | | Sputum | Upregulated | 0.697 (0.616 – 0.779) | sensitivity = 62.67% specificity =78% | 13 | |
| | | - | Upregulated | 0.87 (0.84–0.90) | sensitivity = 82% specificity =62.96% | 12 | |
| 8 | miRNA- 150 | Plasma | Upregulated | 0.752 sensitivity = 81.8% specificity =81.8% | | 8 | |
| 9 | miRNA- let-7a | Serum | Down regulated | 0.847sensitivity = 68.5% specificity =91.1% | | 14 | |

 Table 1: The Relationship between the Expression Levels, Area Under the ROC Curve (AUC) and Diagnostic

 Value of miRNA in Lung Cancer Patients

| S. No | No miRNA Type of sample | | Expression | AUC (95% CI) | Diagnostic value | Ref. No. |
|-------|-------------------------|-------------|-------------------|--|--|----------|
| 10 | miRNA- 375 | Sputum | Upregulated | 0.666 (0.594 – 0.756) | sensitivity = 66.23% specificity =62.22% | 13 |
| | | Plasma | - | 0.6088 (0.4721 - 0.7455) | sensitivity = 66.7% specificity = 57.58% | 27 |
| | | Sputum | Upregulated | 0.713 | sensitivity = 60.3% specificity =71.7% | 16 |
| 11 | miRNA- 21-5p | Plasma | Upregulated | 0.739 (0.670 –0.808) | | 26 |
| | | Plasma | Upregulated | 0.8913 (0.8394 - 0.9431) | sensitivity = 80% specificity =80% | 6 |
| | | Plasma | Upregulated | 0.653 | sensitivity = 79.17% specificity =55.15% | 36 |
| | miRNA- 21 | Sputum | Upregulated | 0.819 (0.753 - 0.874) | sensitivity = 78.16% specificity =71.08% | 13 |
| | | Sputum | Upregulated | 0.752 | sensitivity =58.8 % specificity =73.8% | 16 |
| | | Plasma | Upregulated | 0.5862 (0.4348 - 0.7186) | sensitivity = 56.25% specificity =63.64% | 27 |
| 12 | miRNA- 210 | Plasma | Upregulated | 0.6913 (0.5611 - 0.8215) | sensitivity = 56.25% specificity =72.73% | 27 |
| | | Serum | Upregulated | 0.73 (0.63–0.85) | sensitivity = 76.8% specificity =72.3% | 28 |
| | | Serum | Upregulated | 0.616 (0.534 –0.694) | sensitivity = 33.9% specificity =100% | 29 |
| | | Body fluids | Upregulated | $\begin{array}{c} 0.77 \\ (0.73 - 0.80) \end{array}$ | sensitivity = 65% specificity = 76% | 30 |
| | | Sputum | Upregulated | 0.853 (0.792 –0.901) | sensitivity = 75.27% specificity =85.88% | 13 |
| 13 | miRNA- 486-5p | Plasma | Down regulated | 0.6288 (0.5779 – 0.8312) | sensitivity = 71.8% specificity =66.67% | 27 |
| | | Sputum | Down regulated | $\begin{array}{c} 0.748 \\ (0.674 \ -0.821) \end{array}$ | sensitivity = 74.03% specificity = 66.67% | 13 |
| | miRNA- 486 | Sputum | Down regulated | 0.727 | sensitivity = 62.6% specificity = 69.4% | 16 |
| | | Plasma | Upregulated | 0.926 | sensitivity = 90.9 % specificity = 81.8 % | 8 |
| 14 | miRNA- 708 | Sputum | Down regulated | $\begin{array}{c} 0.656 \\ (0.561 \ -0.750) \end{array}$ | sensitivity = 64.71% specificity =62.50% | 13 |
| 15 | miRNA- 200b | Sputum | Upregulated | 0.679 (0.589 -0.768) | sensitivity = 65.22% specificity =61.19% | 13 |
| | | Sputum | Upregulated | 0.789 | sensitivity = 55.1% specificity =72.2 % | 16 |

 Table 1: The Relationship between the Expression Levels, Area Under the ROC Curve (AUC) and Diagnostic

 Value of miRNA in Lung Cancer Patients

| S. No miRNA Type of sample | | | | AUC (95% CI) | Diagnostic value | Ref. No. | |
|----------------------------|---------------|---------------------|-------------------|--|--|----------|--|
| 16 | miRNA- 182 | Sputum | Upregulated | 0.684 (0.611 - 0.778) | sensitivity = 64.94% specificity =59.76% | 13 | |
| | | Plasma | Upregulated | 0.77 (0.68 – 0.87) | sensitivity = 70%, specificity =79% | 7 | |
| | | Sputum | Upregulated | 0.825 | sensitivity = 64.3%, specificity =79.5% | 16 | |
| | | Serum | Upregulated | 0.734 (0.657 –0.803) | sensitivity = 63.4%, specificity =80% | 29 | |
| | | Tissue | Upregulated | 0.825 | sensitivity = 64.3 %, specificity = 79.5 % | 16 | |
| | | Plasma | Upregulated | 0.7081 (0.6246 –0.7916) | | 25 | |
| 17 | miRNA- 372 | Sputum | Upregulated | 0.707 (0.628 - 0.786) | sensitivity = 63.64%, specificity =60.98% | 13 | |
| 18 | miRNA- 143 | Sputum | Upregulated | 0.723 (0.644 –0.803) | sensitivity = 63.38% specificity = 61.73% | 13 | |
| | | Tissue | Down regulated | 0.97 | sensitivity = 99% specificity = 83% | 35 | |
| 19 | miRNA- 126 | Sputum | Down regulated | $\begin{array}{c} 0.777 \\ (0.704 \ -0.851) \end{array}$ | sensitivity = 77.63% specificity = 75.00% | 13 | |
| | | Serum | Down regulated | 0.793 (0.719 –0.854) | sensitivity = 60.7% specificity = 92.5% | 29 | |
| | | Plasma | Down regulated | 0.5767 (0.4348 – 0.7186) | sensitivity = 62.5% specificity = 63.64% | 27 | |
| | | Sputum | Down regulated | 0.824 | sensitivity = 67.2% specificity = 73.8% | 16 | |
| 20 | miRNA- 31 | Plasma | Upregulated | 0.71 (0.61 – 082) | sensitivity = 73% , specificity = 61% | 7 | |
| | | Peripheral blood | Upregulated | 0.785 (0.486–0.763) | sensitivity = 76.9% specificity = 74.5% | 31 | |
| | | Sputum | Upregulated | 0.789 (0.719–0.849) | sensitivity = 60.23% specificity = 82.67% | 13 | |
| 21 | miRNA- 145 | Sputum | Down regulated | 0.807 | sensitivity = 59.5 %, specificity = 82.9% | 16 | |
| 22 | miRNA- 27a | Plasma | Upregulated | 0.95 (0.9 –0.99) | sensitivity =94%, specificity =81% | 7 | |
| 23 | miRNA- 195 | Plasma | Down regulated | 0.82 (0.74 - 0.90) | sensitivity =74%, specificity =80% | 7 | |
| | | _ | Down regulated | 0.92 | sensitivity = 79%, specificity =100% | 35 | |

Cont. Table 1: The Relationship between the Expression Levels, Area Under the ROC Curve (AUC) and Diagnostic Value of miRNA in Lung Cancer Patients

| S. No miRNA | | Type of sample | Expression | AUC (95% CI) | Diagnostic value | Ref. No. | |
|-------------|-------------------|--|-------------------|---------------------------|--|----------|--|
| 24 | miRNA- 23a | Plasma | Upregulated 0.742 | | sensitivity = 50.00% specificity =92.31% | 36 | |
| 25 | miRNA- 205 | Sputum | Upregulated | 0.635 (0.552 –0.719) | sensitivity = 59.74%, specificity =53.93% | 13 | |
| 26 | miRNA- 30a | Plasma | Upregulated | 0.727 (0.645 0.810) | sensitivity =61%, specificity =84.3% | 33 | |
| 27 | miRNA- 197 | Plasma | Upregulated | 0.8792 (0.8254–0.9330) | | 25 | |
| 28 | miRNA- 661 | Serum | Upregulated | 0.726 | sensitivity = 60.7%, specificity =84.6% | 34 | |
| 29 | miRNA- 181a-5p | Plasma | Upregulated | 0.731 (0.661-0.800) | | 26 | |
| 30 | miRNA- 106a-5p | Plasma | Upregulated | 0.737 (0.667-0.807) | | 26 | |
| 31 | miRNA- 93-5p | Plasma | Upregulated | 0.687 (0.614–0.761) | | 26 | |
| 32 | miRNA- 183 | Serum | Upregulated | 0.626 (0.554-0.703) | sensitivity = 41.1%, specificity =82.5% | 29 | |
| 33 | miRNA- 576-3p | Peripheral blood mononucleated cells | Upregulated | 0.7576 (0.6667–0.8486) | | 15 | |
| 34 | miRNA- 19b-3p | Peripheral blood mononucleated cells | Upregulated | 0.7546 (0.6651-0.8441) | | 15 | |
| 35 | miRNA- 29b-3p | Peripheral blood mononucleated cells | Down regulated | 0.7536 (0.6589–0.8482) | | 15 | |
| 36 | miRNA- 29a-3p | | Down regulated | 0.6050 (0.4969–0.7131) | | 15 | |
| 37 | miRNA- 628-3p | Plasma | Upregulated | 0.730 | sensitivity =42.7%, specificity =91.2% | 32 | |
| 38 | miRNA- 425-3p | Plasma | Upregulated | 0.734 | sensitivity =67.1%, specificity =68.1% | 32 | |
| 39 | miRNA- 532 | Plasma | Down regulated | 0.662 | sensitivity =53.7%, specificity =80.2% | 32 | |
| 40 | miRNA- 339-3p | Plasma | Upregulated | 0.720 | sensitivity =64.6%, specificity =71.4% | 32 | |
| 41 | miRNA- 26b | Plasma | Down regulated | 0.657 (0.616 – 0.696) | | 23 | |
| 42 | miRNA- 19b | Plasma | Down regulated | 0.559 (0.517 – 0.600) | | 23 | |

Cont. Table 1: The Relationship between the Expression Levels, Area Under the ROC Curve (AUC) and Diagnostic Value of miRNA in Lung Cancer Patients

| S.No | Name of miRNA | Age | Gender | Tumor size | Tumor stage | Smoking | Ref. No. |
|------|---------------|--------------|--------------|---------------|----------------|--------------|---------------|
| 1 | miRNA-33a-5p | ✓ | ✓ | ✓ | ✓ | \checkmark | 22 |
| 2 | miRNA-128-3p | ✓ | ✓ | \checkmark | ✓ | \checkmark | 22 |
| 3 | miRNA-17 | ✓ | | | \checkmark | \checkmark | 11 |
| 4 | miRNA-17-5p | ✓ | ✓ | ✓ | ✓ | | 24 |
| 5 | miRNA-155 | ✓ | \checkmark | | \checkmark | \checkmark | 25 |
| 6 | miRNA-197 | ✓ | ✓ | | ✓ | \checkmark | 25 |
| 7 | miRNA-182 | ✓ | ✓ | ✓ | ✓ | \checkmark | 7, 25, 29 |
| 8 | miRNA-183 | | | ✓ | \checkmark | \checkmark | 29 |
| 9 | miRNA-21 | ✓ | ✓ | \checkmark | ✓ | \checkmark | 6, 16, 27, 36 |
| 10 | miRNA-let-7a | ✓ | ✓ | ✓ | ✓ | \checkmark | 14 |
| 11 | miRNA-210 | ✓ | ✓ | \checkmark | \checkmark | \checkmark | 27, 29, 30 |
| 12 | miRNA-31 | ✓ | | \checkmark | ✓ | \checkmark | 7, 31 |
| 13 | miRNA-222 | ✓ | | | \checkmark | \checkmark | 11 |
| 14 | miRNA-19b-3p | ✓ | ✓ | | \checkmark | \checkmark | 15 |
| 15 | miRNA-29b-3p | ✓ | ✓ | | ✓ | \checkmark | 15 |
| 16 | miRNA-628-3p | ✓ | ✓ | | ✓ | \checkmark | 32 |
| 17 | miRNA-425- 3p | ✓ | ✓ | | ~ | \checkmark | 32 |
| 18 | miRNA-532 | ✓ | ✓ | | ✓ | \checkmark | 32 |
| 19 | miRNA-339-3p | ✓ | ✓ | | ✓ | \checkmark | 32 |
| 20 | miRNA-195 | | \checkmark | \checkmark | ✓ | \checkmark | 7 |
| 21 | miRNA-195-5p | | \checkmark | | | \checkmark | 35 |
| 22 | miRNA-23a | ✓ | | \checkmark | ✓ | \checkmark | 36 |
| 23 | miRNA-375 | ✓ | ✓ | \checkmark | \checkmark | \checkmark | 16, 27 |
| 24 | miRNA-486 | ✓ | ✓ | \checkmark | ✓ | \checkmark | 16, 27 |
| 25 | miRNA-200b | \checkmark | ✓ | | ✓ | ✓ | 16 |
| 26 | miRNA-27a | ✓ | ✓ | \checkmark | ✓ | \checkmark | 7 |
| 27 | miRNA-126 | \checkmark | ✓ | ✓ | ✓ | ✓ | 27, 29 |
| 28 | miRNA-30a | ~ | ~ | | ~ | ✓ | 33 |
| 29 | miRNA-661 | ✓ | ✓ | \checkmark | ✓ | | 34 |

 Table 2: Clinical Association of the miRNAs Reported in Lung Cancer Patients

 Significant

 Non-Significant

CONCLUSION

This review article signifies the evaluation of various miRNAs as diagnostic biomarkers in LC patients. As the miRNAs target several genes they may serve as potential therapeutic targets. It may further be beneficial in controlling several other oncogenic signalling cascades.

REFERENCES

- 1. World Health Organization. Global Health Observatory. Geneva: World Health Organization; 2018. gco.iarc.fr/. Accessed September 30, 2020
- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394-424
- 3. Brindle L, Pope C, Corner J, et al. Eliciting symptoms interpreted as normal by patients with early-stage lung cancer: could GP elicitation of normalised symptoms reduce delay in diagnosis? Cross-sectional interview study. BMJ Open. 2012;2(6):e001977
- 4. Levitsky A, Pernemalm M, Bernhardson BM, et al. Early symptoms and sensations as predictors of lung cancer: a machine learning multivariate model. Sci Rep. 2019;9(1):16504
- 5. World Health Organization. Global Health Observatory. Geneva: World Health Organization; 2014. https://gco.iarc.fr/ survival/survmark/Accessed September 30, 2020
- 6. Abu-Duhier FM, Javid J, Sughayer MA, et al. Clinical Significance of Circulatory miRNA-21 as an Efficient Non-Invasive Biomarker for the Screening of Lung Cancer Patients. Asian Pac J Cancer Prev. 2018;19(9):2607-2611
- Szczyrek M, Kuźnar-Kamińska B, Grenda A, et al. Diagnostic value of plasma expression of microRNAs complementary to Drosha and Dicer in lung cancer patients. Eur Rev Med Pharmacol Sci. 2019 May;23(9):3857-3866
- Li W, Wang Y, Zhang Q, et al. MicroRNA-486 as a Biomarker for Early Diagnosis and Recurrence of Non-Small Cell Lung Cancer. PLoS One. 2016;11(1):e0148589
- 9. Obernosterer G, Leuschner PJ, Alenius M, et al. Post-transcriptional regulation of microRNA expression. RNA. 2006;12(7):1161–1167
- 10. Dhawan A, Scott JG, Harris AL, et al. Pan-cancer characterisation of microRNA across cancer hallmarks reveals microRNA-mediated downregulation of tumour suppressors. Nat

Commun. 2018 Dec;9(1):5228

- 11. Hetta HF, Zahran AM, El-Mahdy RI, et al. Assessment of Circulating miRNA-17 and miRNA-222 Expression Profiles as Non-Invasive Biomarkers in Egyptian Patients with Non-Small-Cell Lung Cancer. Asian Pac J Cancer Prev. 2019;20(6):1927-1933
- 12. Shao C, Yang F, Qin Z, et al. The value of miR-155 as a biomarker for the diagnosis and prognosis of lung cancer: a systematic review with metaanalysis. BMC Cancer. 2019 Nov;19(1):1103
- 13. Xing L, Su J, Guarnera MA, et al. Sputum microRNA biomarkers for identifying lung cancer in indeterminate solitary pulmonary nodules. Clin Cancer Res. 2015 Jan 15;21(2):484-489
- 14. Liu JK, Liu HF, Ding Y, et al. Predictive value of microRNA let-7a expression for efficacy and prognosis of radiotherapy in patients with lung cancer brain metastasis: A case–control study. Medicine (Baltimore). 2018 Nov;97(44):e12847
- Ma J, Lin Y, Zhan M, et al. Differential miRNA expressions in peripheral blood mononuclear cells for diagnosis of lung cancer. Lab Invest. 2015 Oct;95(10):1197-1206
- Yu L, Todd NW, Xing L, et al. Early detection of lung adenocarcinoma in sputum by a panel of microRNA markers. Int J Cancer. 2010 Dec;127(12):2870-2878
- 17. Aryani A, Denecke B. In vitro application of ribonucleases: comparison of the effects on mRNA and miRNA stability. BMC Res Notes. 2015;8:164
- Hansen TB, Wiklund ED, Bramsen JB, et al. miRNA-dependent gene silencing involving Ago2-mediated cleavage of a circular antisense RNA. EMBO J. 2011;30(21):4414-4422
- Miyoshi K, Tsukumo H, Nagami T, et al. Slicer function of Drosophila Argonautes and its involvement in RISC formation. Genes Dev. 2005;19(23):2837-2848
- 20. Cambronne XA, Shen R, Auer PL, et al. Capturing microRNA targets using an RNA-induced silencing complex (RISC)-trap approach. Proc Natl Acad Sci U S A. 2012;109(50):20473-20478
- 21. Zhang Y, Li Z, Cheng Y, et al. Colorimetric detection of microRNA and RNase H activity in homogeneous solution with cationic polythiophene derivative. Chem Commun (Camb). 2009 ;(22):3172-3174
- 22. Pan J, Zhou C, Zhao X, et al. A two-miRNA signature (miR-33a-5p and miR-128-3p) in whole

blood as potential biomarker for early diagnosis of lung cancer. Sci Rep. 2018;8(1):16699

- 23. Lua S, Kong H, Hou Y, et al. Two plasma microRNA panels for diagnosis and subtype discrimination of lung cancer. 2018;123:44-51
- 24. Zhang Y, Zhang Y, Yin Y, et al. Detection of circulating exosomal miR-17-5p serves as a novel non-invasive diagnostic marker for non-small cell lung cancer patients. Pathol Res Pract. 2019;215(8):152466
- 25. Zheng D, Haddadin S, Wang Y, et al. Plasma microRNAs as novel biomarkers for early detection of lung cancer. Int J Clin Exp Pathol. 2011;4(6):575–586
- Shan X, Zhang H, Zhang L, et al. Identification of four plasma microRNAs as potential biomarkers in the diagnosis of male lung squamous cell carcinoma patients in China. Cancer Med. 2018 (6): 2370–2381
- 27. Shen J, Liu Z, Todd NW, Zhang H, Liao J, Yu L, Guarnera MA, Li R, Cai L, Zhan M, Jiang F. Diagnosis of lung cancer in individuals with solitary pulmonary nodules by plasma microRNA biomarkers. BMC Cancer. 2011 Aug 24;11:374. doi: 10.1186/1471-2407-11-374.
- Li XX, Liu Y, Meng HH, et al. Expression of miR-210 in senile COPD complicating primary lung cancer. Eur Rev Med Pharmacol Sci . 2017;21(3 Suppl):38-42
- 29. Zhu WY, Zhao KY, Zha Y, Chen DD, et al. Diagnostic Value of Serum miR-182, miR-183, miR-210, and miR-126 Levels in Patients with Early-Stage Non-Small Cell Lung Cancer. PLoS

One. 2016;11(4):e0153046

- 30. He RQ, Cen WL, Cen JM. et al. Clinical Significance of miR-210 and its Prospective Signaling Pathways in Non-Small Cell Lung Cancer: Evidence from Gene Expression Omnibus and the Cancer Genome Atlas Data Mining with 2763 Samples and Validation via Real-Time Quantitative PCR. Cell Physiol Biochem. 2018;46(3):925-952
- 31. Yan HJ, Ma JY, Wang L, et al. Expression and significance of circulating microRNA-31 in lung cancer patients. Med Sci Monit. 2015;21:722-726
- 32. Wang Y, Zhao H, Gao X, et al. Identification of a three-miRNA signature as a blood-borne diagnostic marker for early diagnosis of lung a d e n o c a r c i n o m a . O n c o t a r g e t . 2016;7(18):26070–26086
- 33. Sun L, Chen Y, Su Q, et al. Increased Plasma miRNA-30a as a Biomarker for Non-Small Cell Lung Cancer. Med Sci Monit. 2016;22:647–655
- 34. Zhou GH, Yang WH, Sun B. Clinical impact of serum miR-661 in diagnosis and prognosis of non-small cell lung cancer. Eur Rev Med Pharmacol Scisa. 2017;21(24):5696-5701
- 35. Li L, Feng T, Zhang T, et al. MicroRNA Biomarker hsa-miR-195-5p for Detecting the Risk of Lung Cancer. Int J Genomics . 2020;2020:7415909
- 36. Hetta HF, Zahran AM, Shafik EA, et al. Circulating miRNA-21 and miRNA-23a Expression Signature as Potential Biomarkers for Early Detection of Non-Small-Cell Lung Cancer. Microrna. 2019;8(3):206-215

Orcid ID:

Omair Ahmad - https://orcid.org/0000-0001-9988-240X

Bilal Khan - https://orcid.org/0000-0002-2992-3875

Mohammad Armoghan - https://orcid.org/0000-0002-6598-021X

Swapnil Raj Gautam - https://orcid.org/0000-0003-2676-8010

Zakiya Saiyyada - https://orcid.org/0000-0001-7396-4757

Zainab Siddiqui - https://orcid.org/0000-0003-4384-197X

How to cite this article:

Ahmad O., Saiyada Z, Armoghan M., Khan B, Gautam S. R., Siddiqui Z. Differential expression of microRNAs in lung cancer: A review article. Era J. Med. Res. 2022;9(1):83-91.

Licencing Information

Attribution-ShareAlike 2.0 Generic (CC BY-SA 2.0) Derived from the licencing format of creative commons & creative commonsmay be contacted at https://creativecommons.org/ for further details.