

STRUCTURAL CHARACTERISTICS AND FUNCTIONAL INSIGHTS INTO TSP-4: IMPLICATIONS FOR CARDIOVASCULAR HEALTH

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ABSTRACT

Thrombospondin-4 (TSP-4) is a multidomain protein with unique functions within the thrombospondin family. It plays a works in tissue repair, cell-to-matrix conveying, and various physiological processes. TSP-4 differs structurally from other family members and is associated with a common single nucleotide polymorphism (SNP), A387P, linked to cardiovascular disorders. Its functions include tissue remodeling, regeneration, proliferation, adhesion, migration, angiogenesis, inflammation, and adipogenesis. The A387P SNP in TSP-4 increases the risk of these cardiovascular conditions and affects angiogenesis, inflammation, and adipogenesis. TSP-4 has clinical applications as a marker for various tissues and pathological disorders. Its expression levels differentiate cell origins, making it a biomarker for articular cartilage, tendon progenitor cells, and more. In heart-related conditions, TSP-4 serves as a marker for cardiac overload and coronary artery disease. Additionally, TSP-4 expression is associated with osteoarthritis severity, suggesting its potential as a biomarker for this condition. Overall, understanding TSP-4's diverse functions and its role in cardiovascular pathology provides insights into its clinical relevance and diagnostic potential, making it a promising target for further research and therapeutic interventions.

KEYWORDS: Thrombospondin-4 (TSP-4), Matricellular glycoprotein, Tissue repair, Cell-to-matrix communication, Cardiovascular disorders, Single nucleotide polymorphism (SNP), A387P, Tissue remodeling.

INTRODUCTION

A member of the TSP family, thrombospondin-4 (TSP-4) is a protein that is created by the human Thbs4 gene. TSP-4 performs various roles that are significantly different from those of other family members, and it even competes with them functionally. TSP-4, for instance, has the ability to stimulate angiogenesis (4), whereas TSP-1 and TSP-2 are anti-angiogenic (5). The actions of basement membranes in different tissues are regulated by TSP-4. TSP-4 has been identified in adult hearts (6), muscles of the skeleton (7), articular cartilage cells (8), and it builds up at the neuromuscular junction (9). TSP-4 is a crucial regulator of cell proliferation and remodeling, according to existing research.

Cardiac remodeling serves as a clinical indicator of genomic expression changes influenced by various factors such as cardiac strain, damage, hemodynamic load, and neurohormone activation. These factors contribute to modifications at the molecular, cellular, and extracellular levels, ultimately affecting heart size (10). The progression of heart failure (HF) is commonly attributed to cardiac remodeling (11) (12). Moreover, fluctuations in TSP-4 expression are

frequently observed in conjunction with cardiovascular conditions like myocardial infarction (MI), heart failure, hypertension, arteriosclerosis.

In this review, we highlight the typical function of TSP-4, particularly in relation to heart disease. TSP-1 and TSP-2 are homotrimers in the TSP family, whereas TSP-3, TSP-4, and TSP-5 are homopentamers in subgroup B (13). TSP-4 is a big central particle that can be detected in electron microscopy. It is made up of five subunits that are joined together by globular domains that are located near to the N-terminal (14). The appearance of TSP-4 subunits bears a resemblance to TSP-1 subunits; however, a notable difference lies in the smaller size of TSP-4 subunits (15-18). Notably, heparin is present in TSP-5 but not in TSP-4 (19). Heparin affinity chromatography allows for the enrichment of TSP-4 (20).

A prevalent single nucleotide polymorphism (SNP) linked to cardiovascular disorders within TSP-4 is A387P (22). This specific SNP substitution takes place in the third continuous type III, and it is recognized that this variant has the potential to enhance calcium binding (23).

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Function

The majority of TSP-4's actions are intercellular because it is an intercellular protein. TSP-4 controls the extracellular matrix (ECM) composition in the majority of its enriched locations, which include the muscles and tendons. The fibril formation to the surface of cells may be mediated by TSP-4 (24). Additionally, TSP-4 deficiency alters the structure and physiological operation of tissues (25) and is a significant contributor to cardiovascular disease. Previous research has shown that TSP-4 is essential for migration, attachment, remodeling, recuperation, and inflammation as well as for promoting growth of the nervous system (26).

Renovation and regrowth

The TSP family, whose production is frequently stimulated, supports tissue regeneration through stress (27). The first and most frequently described function of TSP-4 is tissue remodeling and regeneration. During tissue injury, regeneration, and renovation, TSP-4 expression is frequently unregulated (28). TSP-4 is implicated in the remodeling process because it has the ability to change the composition of the matrix proteins (29). TSP-4 binds to and controls structural ECM proteins in heart cellstroma during tissue injury (30), which encourages adaptive ECM remodeling. Additionally, cardiac fibrosis is controlled by cardiomyocytes by transcriptionally modulating TSP-4, which in turn controls cardiac fibroblast activation (31). Additionally, through the Toll-like receptor 4 signaling system, TSP-4 can increase the production of the elastin gene and myofibroblast alteration, resulting in the fibrosis of the epidermis (32). In vivo limb regeneration is influenced by TSP (33). Additionally, the overexpression of TSP-4 induces non-hepatocytes in the vicinity of the injured liver's portal region to support liver regeneration (34).

Proliferation

Cell proliferation stands out as a fundamental function of TSP-4, particularly within the cardiovascular system, akin to activities such as tissue remodeling and tissue regeneration. The modulation of endothelial cell is crucial for vessel wall dynamics, is under the control of vascular cells producing TSP-4 (35). Through interactions with integrin 2 and gabapentin receptor 21, recombinant TSP-4 expedites the development of vascular smooth muscle cells (36) and microvascular endothelial cells (37), while significantly impeding the growth of endothelial cells derived from TSP-4 knockout mice (38). The efficacy of TSP-4 in promoting endothelial cell proliferation undergoes substantial changes due to the single nucleotide polymorphism (SNP) in TSP-4 (39). Furthermore, the C-terminal peptide of TSP-4 (C21) exhibits the ability

to enhance the proliferation of erythroid cells (40). C21 amplifies thymidine erythrocytes and fibroblasts from bovine skin (41).

Adhesion and migration

TSP-4 is implicated in cell adhesion and migration, according to numerous studies (42). TSP-4 is necessary for cellular adhesion (43). In the ECM, TSP-4 controls molecules that promote axonal development and adhesion (44). Fusion proteins including C21 promote myoblast adhesion, according to earlier research (26). (45). The ability of TSP-4 to facilitate endothelial cell adhesion is significantly altered by an SNP mutation (39). Additionally, it has been noted that suppressing the lncRNA TSP-4 prevents prostate cancer cells from migrating and invading (46).

Angiogenesis

Angiogenesis is a crucial process for growth and development, and the production of granulation tissue. The same time numerous endothelial cell-related responses are impacted by TSP-4, including the development of increased angiogenesis (37). For instance, TSP-4 enhances TGF-1's effect on angiogenesis (38). TSP-4 also improves endothelial cells' capacity to stimulate angiogenesis, which aids in the promotion of neovascularization (47). According to studies, TSP-4 knockdown and knockout (KO) suppress the angiogenesis caused by hepatocellular cancer (48) and TSP-4 KO mice have less angiogenesis (49).

Inflammation

The biological process of inflammation shields the body from damaging stimuli. TSP-4 has a crucial role in controlling vascular inflammation (50). Inflammation considerably increases the expression of TSP-4 (28). A glycoprotein implicated in the inflammatory response is encoded by *Thbs4*. The inflammatory response to neuropathic pain is mediated by TSP-4 (51). Interleukin-8 and extra H₂O₂ are released by cells with the TSP-4 A387P mutation (52). Furthermore, TSP-4 suppression lowers other markers of arterial wall inflammation, inhibits endothelial cell activation, and greatly reduces the amount of macrophages in the lesion (45).

Adipogenesis

Pre-adipocytes differentiate into adipocytes through a process known as adipogenesis. In mice, TSP-4 is a putative myokine that is related with obesity and exercise (53). When compared to mature adipocytes, TSP-4 is upregulated in human pre-adipocytes (54). TSP-4 may therefore play a role in the early stages of adipogenesis. TSP-4 also facilitates the contact between muscle fibers and ECM (7).

Vascular conditions

Cardiovascular pathology is significantly impacted by TSPs and associated SNPs (50). It's interesting to note that whereas the N700S SNP in TSP-1 decreases Ca²⁺ binding production, the A387P SNP in TSP-4 increases Ca²⁺ binding (23). Cardiovascular illness is brought on by both TSP mutations, despite their divergent mechanisms.

Myocardial infarction

Research has demonstrated that TSP-4 promotes ER stress and specifically inhibits TGF- in myocytes, acting as a protective mechanism for cardiomyopathy and lowering early mortality after MI (55).

Following myocardial infarction (MI) in rats, elevated TSP-4 mRNA levels were observed (56), and a noteworthy correlation between the A387P variation of TSP-4 and MI was identified (57). Subsequent investigations revealed that the TSP-4 A387P variation plays a crucial role in the onset of MI across all age groups (58). Divergent research findings also exist. In one study, the TSP-4 A387P polymorphism, specifically in the female homozygous state, was exclusively associated with MI in females (22), whereas another study report homozygosity of the TSP-4 A387P variant to elderly women (59). Two separate studies underscore the substantial and independent association of the TSP-4 A387P genotype with MI risk in both American and Egyptian populations (60, 61).

Enlarged ventricles and cardiac failure

After hypertensive cardiac disease, TSP-4 is linked to myocardial remodeling (62), which may result in ventricular hypertrophy. In addition, abnormal ventricular hypertrophy frequently results in ventricular dilatation, which impairs the heart's ability to efficiently pump blood and causes HF. Additionally, TSP-4 is crucial in controlling HF remodeling (63). Thbs4(-/-) exhibits severe weaknesses in coping with chronic stress overload, according to animal investigations. Additionally, mice with end-stage dilated cardiomyopathy (6), monocrotaline-induced pulmonary hypertension (64,65) were found to have TSP-4 overexpression.

Hypertension

Persistent hypertension stands out as a prominent factor for various other cardiovascular ailments. Within the context of hypertension, the regulation of cardiac hypertrophy progression, influence on aortic aneurysm formation, and modification of endothelium-dependent resistance to arterial relaxation are orchestrated by TSP-4 (11).

Coronary artery disease

A decrease in the CAD, is brought on by a number of reasons. In the beginning, a study in the US population discovered a strong connection between CAD and the TSP-4 A387P SNP (66). More studies later supported this discovery and demonstrated that the A387P SNP of TSP-4 is linked to an elevated risk of CAD in the American population (67). In contrast, one study (9) found no connection between the TSP-4 A387P mutation and CAD in the target group.

Peripheral artery disease

At initially, atherosclerosis typically exhibits no symptoms, but in extreme circumstances, it can cause renal issues, CAD, stroke, and peripheral artery disease. TSP-4 is implicated in the formation of atherosclerosis and is concentrated in atherosclerotic lesions and regions prone to pathogenesis (68). As previously indicated, TSP-4 influences endothelial cells, which in turn modulates macrophage recruiting (45). The TSP-4 A387P SNP significantly alters the shape and prevents endothelial cells from adhering to one another and proliferating (35). The mutation is compatible with the phenotype that causes thrombosis and atherosclerosis (39), which suggests that atherosclerosis may be the result of the mutation. One of curcumin's potential mechanisms for preventing a low-density lipoprotein-induced decline in TSP is that it possesses anti-atherosclerotic properties. (69). On the other hand, in one investigation, atherosclerotic lesions showed a drop in TSP-4 levels (70).

Clinical benefit

The discernment of different cell origins can be achieved by assessing TSP-4 expression levels in tissues. TSP-4 serves as a distinctive marker for identifying articular cartilage (8) and distinguishing tendon progenitor cells from peritenon progenitors (71). TSP-4 is expressed at higher levels in SVZ astrocytes in mice than it is in cortical astrocytes (72). Additionally, TSP-4's differential regulation may be used as an biomarker for triggering signal transduction (73).

Specific pathological disorders and diseases can be recognized and diagnosed using TSP-4. TSP-4 is a recognized CAD marker (74) and an endothelial-specific marker for cardiac overload (10). Currently, osteoarthritis can be diagnosed with the support of the finding of IgG isotype autoantibodies against TSP-4 (76). TSP-4 has also been mentioned as a possible biomarker for locoism (77) and the myopathy brought on by a lack of S-adenosylhomocysteine hydrolase (78). Similar to this, TSP-4 and the degree of methylation in it can be useful tumor indicators.

CONCLUSION

In conclusion, thrombospondin-4 (TSP-4) emerges as a multifaceted and pivotal player in various physiological and pathological processes, with a significant impact on cardiovascular health. TSP-4 distinguishes itself by exerting distinct functions compared to other family members, playing a crucial role in tissue remodeling, regeneration, angiogenesis, inflammation, adipogenesis, and cell proliferation. The intricate structure of TSP-4, characterized by homopentameric assembly and unique domains, underscores its versatility in mediating cell-matrix interactions and influencing diverse cellular activities. Notably, TSP-4's involvement in cardiovascular conditions, such as cardiac failure, hypertension highlights its significance in maintaining cardiovascular homeostasis. The impact of single nucleotide polymorphisms (SNPs), particularly the A387P variant, underscores the genetic variability associated with TSP-4 and its role in cardiovascular disorders. This genetic variation can influence calcium binding and contribute to the pathogenesis of conditions like myocardial infarction, enlarging ventricles, and hypertension. Beyond its role in cardiovascular diseases, TSP-4 serves as a valuable marker in various clinical contexts, aiding in the diagnosis and understanding of diseases such as osteoarthritis, atherosclerosis, and myopathy.

REFERENCES

- Arber S, Caroni P. Thrombospondin-4, an extracellular matrix protein expressed in the developing and adult nervous system promotes neurite outgrowth. *J Cell Biol.* 1995; 131: 1083-1094.
- Kazerounian S, Yee KO, Lawler J. Thrombospondins in cancer. *Cell Mol Life Sci.* 2008; 65: 700.
- Lawler J. Thrombospondin-1 as an endogenous inhibitor of angiogenesis and tumor growth. *J Cell Mol Med.* 2002; 6: 1-12.
- Muppala S, Frolova E, Xiao R, et al. Proangiogenic properties of thrombospondin-4. *arterioscler. Thromb Vasc Biol.* 2015; 35: 1975-1986.
- Lawler J. The functions of thrombospondin-1 and -2. *Curr Opin Cell Biol.* 2000; 12: 634-640
- Tan FL, Moravec CS, Li J, et al. The gene expression fingerprint of human heart failure. *Proc Natl Acad Sci U S A.* 2002; 99: 11387- 11392.
- Albrecht E, Schering L, Liu Y, et al. Triennial growth and development symposium: factors influencing bovine intramuscular adipose tissue development and cellularity. *J Anim Sci.* 2017; 95: 2244-2254.
- Jeschke A, Bonitz M, Simon M, et al. Deficiency of thrombospondin-4 in mice does not affect skeletal growth or bone mass acquisition, but causes a transient reduction of articular cartilage thickness. *PLoS One.* 2015; 10: e0144272.
- Kasprick DS, Kish PE, Junttila TL, et al. Microanatomy of adult zebrafish extraocular muscles. *PLoS One.* 2011; 6: e27095.
- Mustonen E, Aro J, Puhakka J, et al. Thrombospondin-4 expression is rapidly upregulated by cardiac overload. *Biochem Biophys Res Commun.* 2008; 373: 186-191.
- Cohn JN, Ferrari R, Sharpe N. Cardiac remodeling--concepts and clinical implications: a consensus paper from an international forum on cardiac remodeling. Behalf of an International Forum on Cardiac Remodeling. *J Am Coll Cardiol.* 2000; 35: 569-582.
- Palao T, Rippe C, van Veen H, et al. Thrombospondin-4 knockout in hypertension protects small-artery endothelial function but induces aortic aneurysms. *Am J Physiol Heart Circ Physiol.* 2016; 310: H1486-H1493.
- Lawler J, McHenry K, Duquette M, et al. Characterization of human thrombospondin-4. *J Biol Chem.* 1995; 270: 2809-2814.
- Narouz-Ott L, Maurer P, Nitsche DP, et al. Thrombospondin-4 binds specifically to both collagenous and non-collagenous extracellular matrix proteins via its C-terminal domains. *J Biol Chem.* 2000; 275: 37110-37117.
- Mörgelin M, Heinegård D, Engel J, et al. Electron microscopy of native cartilage oligomeric matrix protein purified from the Swarm rat chondrosarcoma reveals a fivearmed structure. *J Biol Chem.* 1992; 267: 6137-6141.
- Selander-Sunnerhagen M, Ullner M, Persson E, et al. How an epidermal growth factor (EGF)-like domain binds calcium. High resolution NMR structure of the calcium form of the NH₂-terminal EGF-like domain in coagulation factor X. *J Biol Chem.* 1992; 267: 19462-19469.
- Schips TG, Vanhoutte D, Vo A, et al. Thrombospondin-3 augments injury-induced cardiomyopathy by intracellular integrin inhibition and sarcolemmal instability. *Nat Commun.* 2019; 10: 76.
- Misenheimer TM and Mosher DF. Biophysical characterization of the signature domains of thrombospondin-4 and thrombospondin-2. *J Biol Chem.* 2005; 280: 41229-41235.

19. Park JF, Yu YP, Gong N, et al. The EGF-LIKE domain of thrombospondin-4 is a key determinant in the development of pain states due to increased excitatory synaptogenesis. *J BiolChem.* 2018;293: 16453-16463.
20. Hauser N, Paulsson M, Kale AA, et al. Tendon extracellular matrix contains pentameric thrombospondin-4 (TSP-4). *FEBS Lett.* 1995; 368: 370-310.
21. Södersten F, Ekman S, Schmitz M, et al. Thrombospondin-4 and cartilage oligomeric matrix protein form heterooligomers in equine tendon. *Connect Tissue Res.* 2006; 27: 85-91.
22. Cui J, Randell E, Renouf J, et al. Gender dependent association of thrombospondin-4 A387P polymorphism with myocardial infarction. *Arterioscler.ThrombVascBiol.* 2004; 24: e183- e184.
23. Stenina OI, Ustinov V, Krukovets I, et al. Polymorphisms A387P in thrombospondin-4 and N700S in thrombospondin-1 perturb calcium binding sites. *FASEB J.* 2005; 19: 1893-1895.
24. Södersten F, Ekman S, Niehoff A, et al. Ultrastructural immunolocalization of cartilage oligomeric matrix protein, thrombospondin-4, and collagen fibril size in rodent achilles tendon in relation to exercise. *Connect Tissue Res.* 2007; 48: 254-262.
25. Frolova EG, Drazba J, Krukovets I, et al. Control of organization and function of muscle and tendon by thrombospondin-4. *Matrix Biol.* 2014; 37: 35-48.
26. Adams JC, Lawler J. Cell-type specific adhesive interactions of skeletal myoblasts with thrombospondin-1. *MolBiol. Cell* 1994; 5: 423- 437.
27. Brody MJ, Schips TG, Vanhoutte D, et al. Dissection of thrombospondin-4 domains involved in intracellular adaptive endoplasmic reticulum stress-responsive signaling. *Mol Cell Biol.* 2016; 36: 2-12.
28. Niu J, Lin Y, Liu P, et al. Microarray analysis on the lncRNA expression profile in male hepatocellular carcinoma patients with chronic hepatitis B virus infection. *Oncotarget.* 2016; 7: 76169-76180.
29. De Groote MA, Sterling DG, Hraha T, et al. Discovery and validation of a six-marker serum protein signature for the diagnosis of active pulmonary tuberculosis. *J ClinMicrobiol.* 2017; 55: 3057-3071.
30. Doroudgar S and Glembotski CC. ATF6 (corrected) and thrombospondin 4: the dynamic duo of the adaptive endoplasmic reticulum stress response. *Circ Res.* 2013; 112: 9-12.
31. Sawaki D, Hou L, Tomida S, et al. Modulation of cardiac fibrosis by Krüppel-like factor 6 through transcriptional control of thrombospondin 4 in cardiomyocytes. *Cardiovasc Res.* 2015; 107: 420- 430.
32. Moon SJ, Bae JM, Park KS, et al. Compendium of skin molecular signatures identifies key pathological features asso- The role of thrombospondin-4 in cardiovascular diseases 367 *Int J ClinExp Med.* 2020;13(2):358-370.
33. Whited JL, Lehoczky JA and Tabin CJ. Inducible genetic system for the axolotl. *Proc. Natl. Acad. Sci.* 2012; 109: 13662-13667.
34. Klaas M, Kangur T, Viil J, et al. The alterations in the extracellular matrix composition guide the repair of damaged liver tissue. *Sci. Rep.* 2016; 6: 27398.
35. Stenina OI, Desai SY, Krukovets I, et al. Thrombospondin-4 and its variants: expression and differential effects on endothelial cells. *Circulation.* 2003; 108: 1514-1519.
36. Lv L, Liang W, Ye M, et al. Thrombospondin-4 ablation reduces macrophage recruitment in adipose tissue and neointima and suppresses injury-induced restenosis in mice. *Atherosclerosis.* 2016; 247: 70-77.
37. Muppala S, Frolova E, Xiao R, et al. Proangiogenic properties of thrombospondin-4. *arterioscler. Thromb Vasc Biol.* 2015; 35: 1975-1986.
38. Muppala S, Xiao R, Krukovets I, et al. Thrombospondin-4 mediates TGF- β -induced angiogenesis. *Oncogene.* 2017; 36: 5189-5198.
39. Stenina OI, Byzova TV, Adams JC, et al. Coronary artery disease and the thrombospondin single nucleotide polymorphisms. *Int J Biochem Cell Biol.* 2004; 36: 1013-1030.
40. Sadvakassova G, Dobocan MC, Difalco MR, et al. Regulator of differentiation 1 (ROD1) binds to the amphipathic C-terminal peptide of thrombospondin-4 and is involved in its mitogenic activity. *J Cell Physiol.* 2009; 220: 672-679.
41. Congote LF, Difalco MR and Gibbs BF. The Cterminal peptide of thrombospondin-4 stimulates erythroid cell proliferation. *Biochem Biophys Res Commun.* 2004; 324: 673-678.
42. Kondo Y, Shen L, Yan PS, et al. Chromatin immunoprecipitation microarrays for identification of genes silenced by histone H3 lysine 9 methylation. *Proc Natl Acad Sci U S A.* 2004; 101: 7398-7403.

43. Ponda MP and Breslow JL. Serum stimulation of CCR7 chemotaxis due to coagulation factor XIIIa-dependent production of high-molecular weight kininogen domain 5. *Proc Natl Acad Sci. U S A.* 2016; 113: E7059-E7068.
44. Dunkle ET, Zaucke F and Clegg DO. Thrombospondin-4 and matrix three-dimensionality in axon outgrowth and adhesion in the developing retina. *Exp Eye Res.* 2007; 84: 707-717.
45. Frolova EG, Pluskota E, Krukovets I, et al. Thrombospondin-4 regulates vascular inflammation and atherogenesis. *Circ Res.* 2010; 107: 1313-1325.
46. Liu J, Cheng G, Yang H, et al. Reciprocal regulation of long noncoding RNAs THBS4-003 and THBS4 control migration and invasion in prostate cancer cell lines. *Mol Med Rep.* 2016; 14: 1451-1458.
47. Zhang Q, Zhou M, Wu X, et al. Promoting therapeutic angiogenesis of focal cerebral ischemia using thrombospondin-4 (TSP4) gene-modified bone marrow stromal cells (BMSCs) in a rat model. *J Transl Med.* 2019; 17: 111.
48. Su F, Zhao J, Qin S, et al. Over-expression of Thrombospondin 4 correlates with loss of miR-142 and contributes to migration and vascular invasion of advanced hepatocellular carcinoma. *Oncotarget.* 2017; 8: 23277-23288.
49. Sure VN and Katakam PV. Janus face of thrombospondin-4: impairs small artery vasodilation but protects against cardiac hypertrophy and aortic aneurysm formation. *Am J Physiol Heart Circ Physiol.* 2016; 310: H1383-H1384.
50. Wilsgaard T, Mathiesen EB, Patwardhan A, et al. Clinically significant novel biomarkers for prediction of first ever myocardial infarction: the Tromsø Study. *Circ Cardiovasc Genet.* 2015; 8: 363-371.
51. Kuttapitiya A, Assi L, Laing K, et al. Microarray analysis of bone marrow lesions in osteoarthritis demonstrates upregulation of genes implicated in osteochondral turnover, neurogenesis and inflammation. *Ann Rheum Dis.* 2017; 76: 1764-1773.
52. Pluskota E, Stenina OI, Krukovets I, et al. Mechanism and effect of thrombospondin-4 polymorphisms on neutrophil function. *Blood* 2005; 106: 3970-3978.
53. Schering L, Hoene M, Kanzleiter T, et al. The role of thrombospondin-4 in cardiovascular diseases 368 *Int J ClinExp Med* 2020;13(2):358-370.
54. Urs S, Smith C, Campbell B, et al. Gene expression profiling in human preadipocytes and adipocytes by microarray analysis. *J Nutr.* 2004; 134: 762-70.
55. Rainer PP, Hao S, Vanhoutte D, et al. Cardiomyocyte-specific transforming growth factor β suppression blocks neutrophil infiltration, augments multiple cytoprotective cascades, and reduces early mortality after myocardial infarction. *Circ Res.* 2014; 114: 1246-1257.
56. Jin H, Yang R, Awad TA, et al. Effects of early angiotensin-converting enzyme inhibition on cardiac gene expression after acute myocardial infarction. *Circulation.* 2001; 103: 736-742.
57. Topol EJ, McCarthy J, Gabriel S, et al. Single nucleotide polymorphisms in multiple novel thrombospondin genes may be associated with familial premature myocardial infarction. *Circulation.* 2001; 104: 2641-2644.
58. Hamsten A and Eriksson P. Thrombospondins and premature coronary artery disease: time to go beyond genotype-phenotype association studies. *Arterioscler. Thromb Vasc Biol.* 2003; 23: 6-7.
59. McCarthy JJ, Parker A, Salem R, et al. Gene Quest Investigators. Large scale associa- The role of thrombospondin-4 in cardiovascular diseases 369 *Int J ClinExp Med* 2020;13(2):358-370.
60. Ye H, Zhou A, Hong Q, et al. Association of seven thrombotic pathway gene CpGSNPs with coronary heart disease. *Biomed Pharmacother.* 2015; 72: 98-102.
61. Abdelmonem NA, Turkey NO, Hashad IM, et al. Association of Thrombospondin-1 (N700S) and Thrombospondin-4 (A387P) gene polymorphisms with the incidence of acute myocardial infarction in egyptians. *Curr Pharm Biotechnol.* 2017; 18: 1078-1087.
62. Mustonen E, Leskinen H, Aro J, et al. Metoprolol treatment lowers thrombospondin-4 expression in rats with myocardial infarction and left ventricular hypertrophy. *Basic Clin Pharmacol Toxicol.* 2010; 197: 709-717.
63. Rysä J, Leskinen H, Ilves M, et al. Distinct upregulation of extracellular matrix genes in transition from hypertrophy to hypertensive heart failure. *Hypertension.* 2005; 45: 927-933.
64. Imoto K, Okada M and Yamawaki H. Expression profile of matricellular proteins in hypertrophied right ventricle of monocrotaline-induced pulmonary hypertensive rats. *J Vet Med Sci.* 2017; 79: 1096-1102.
65. Melenovsky V, Benes J, Skaroupkova P, et al. Metabolic characterization of volume overload

- heart failure due to aorto-caval fistula in rats. *Mol Cell Biochem.* 2011; 354: 83-96.
66. Corsetti JP, Ryan D, Moss AJ, et al. Thrombospondin-4 polymorphism (A387P) predicts cardiovascular risk in postinfarction patients with high HDL cholesterol and C-reactive protein levels. *ThrombHaemost* 2011; 106: 1170-1178.
67. Corsetti JP, Salzman P, Ryan D, et al. Plasminogen activator inhibitor-2 polymorphism associates with recurrent coronary event risk in patients with high HDL and C-reactive protein levels. *PLoS One.* 2013; 8: e68920.
68. Aissa AF, Amaral CLD, Venancio VP, et al. Methionine-supplemented diet affects the expression of cardiovascular disease-related genes and increases inflammatory cytokines in mice heart and liver. *J Toxicol Environ Health Part A.* 2017; 80: 1116-1128.
69. Zhou ZY, Chen YQ, Wang FY, et al. Effect of curcumin on down-expression of thrombospondin-4 induced by oxidized lowdensity lipoprotein in mouse macrophages. *Biomed Mater Eng.* 2014; 24: 181-189.
70. Kristensen LP, Larsen MR, Mickley H, et al. Plasma proteome profiling of atherosclerotic disease manifestations reveals elevated levels of the cytoskeletal protein vinculin. *J Proteomics.* 2014; 101: 141-153.
71. Mienaltowski MJ, Cánovas A, Fates VA, et al. Transcriptome profiles of isolated murine Achilles tendon proper- and peritenon-derived progenitor cells. *J Orthop Res.* 2019; 37: 1409-1418.
72. Zierfuss B, Höbaus C, Herz CT, et al. Thrombospondin-4 increases with the severity of peripheral arterial disease and is associated with diabetes. *Heart Vessels.* 2020; 35: 52-58.
73. Berasi SP, Varadarajan U, Archambault J, et al. Divergent activities of osteogenic BMP2, and tenogenic BMP12 and BMP13 independent of receptor binding affinities. *Growth Factors.* 2011; 29: 128-139.
74. Jing L, Parker CE, Seo D, et al. Discovery of biomarker candidates for coronary artery disease from an APOE-knock out mouse model using iTRAQ-based multiplex quantitative proteomics. *Proteomics.* 2011; 11: 2763-2776.
75. Ruthard J, Hermes G, Hartmann U, et al. The role of thrombospondin-4 in cardiovascular diseases 370 *Int J ClinExp Med* 2020;13(2):358-370
76. Diegl S, Zaucke F, Wagener R, et al. Identification of antibodies against extracellular matrix proteins in human osteoarthritis. *BiochemBiophys Res Commun.* 2018; 503: 1273-1277.
77. Jiao J, Wang S, Zhang R, et al. iTRAQ-based quantitative proteomics discovering potential serum biomarkers in locoweed poisoned rabbits. *Chem Biol Interact.* 2017; 268: 111-118.
78. Sedic M, Kraljevic Pavelic S, Cindric M, et al. Plasma biomarker identification in S-adenosylhomocysteine hydrolase deficiency. *Electrophoresis.* 2011; 32: 1970-1975.

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